



National Institute for Public Health
and the Environment
Ministry of Health, Welfare and Sport

A first analysis of the sensitivity and specificity of seven rapid SARS-CoV-2 antibody tests in clinical patients, including analysis of correlation with neutralization and the Wantai ELISA.

At the end of 2019, SARS-CoV-2 emerged in the human population. The subsequent growing pandemic spread of the virus is accompanied by high morbidity and mortality, and has an enormous negative impact on societal and economic circumstances world-wide. In response to this outbreak and in the context of world-wide shortages for molecular testing, rapid diagnostic tests for detection of SARS-CoV-2 specific antibodies are currently overflowing the diagnostic market. As at 09 April 2020, the FIND organization has listed 155 rapid immuno-assays in different stages of validation and regulation on its website. The added value of these rapid immuno-assays for individual patient diagnostics and their usefulness for epidemiological studies and to direct mitigation strategies, urgently needs to be established.

Here, we took a first look at the clinical sensitivity and specificity of seven rapid SARS-CoV-2 antibody tests and compared the results with the outcomes of a virus neutralization test and a commercial ELISA (Wantai) with apparent high specificity and sensitivity (data not shown).

Methods.

Seven rapid SARS-CoV-2 antibody tests were included in the study. Selection was partially based on pre-study dossier analysis of data provided by the manufacturers that included test-specifics (antigen used), validation data on sensitivity and specificity in relation to type of cohort used and reliability of the manufacturer. Additionally, tests were included that were delivered to the RIVM directly (without solicitation) and that were not triaged based on manufacturer's dossier. The following 7 tests were analyzed in this study:

Manufacturer
1
2
3
4
5 Biomerica
6
7

RIVM

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The following 50 sera were used for the initial screening of the usefulness of the seven rapid immuno-assays:

specificity panel*	number
healthy blood donors (the Netherlands)	10
acute EBV (the Netherlands)	5
acute CMV (the Netherlands)	5
HCoV-OC43 (convalescent, the Netherlands)	5
Total	25
Sensitivity panel*	
acute hospitalized patients (PCR-confirmed)	8
convalescent hospitalized patients (PCR-confirmed)	6
mild illness hospital workers (PCR-confirmed)	11
Total	25

*due to the limited amount of tests available, the validation in this report is limited. Therefore, the data can be used only as a first screening of the utility of the tests. Tests with good performance can be selected for further, more in-depth validation once tests are available.

Furthermore the total number of validation sera used for the different tests varies between 38 and 50.

All tests were used according to manufacturer's instructions. Sera from confirmed SARS-CoV-2 patients were provided by (10)(20) (10)(20), EZT. CMV and EBV acute sera were provided by (10)(20) (10)(20) (LUMC), HCoV-OC43 convalescent sera were provided by (10)(20) 10(120) (RIVM). Serum from healthy blood donors were obtained through Sanquin.

The Wantai ELISA as reference test was performed according to manufacturer's instructions. Virus neutralization test is an in-house test based on a Dutch SARS-CoV-2 isolate (unpublished).

Results.

The seven rapid immuno-assay tests were analyzed for an initial impression of sensitivity and specificity based on clinical samples from confirmed COVID-19 patients and from EBV/CMV/HCOV-OC43 patients/healthy individuals collected before 2019. In tables 1-7 the calculated specifics are depicted per test and are based on PCR-positivity as reference test (Corman et al., 2020).

Tables 1-7. Clinical sensitivity and specificity (%) for seven rapid immuno-assays.

1

A. IgG

Positive/ total no. samples	Cohort	Post onset symptoms (days)	Sensitivity	Specificity
2/8	Acute hospitalized patients	<10	25%	na
6/6	Convalescent hospitalized patients	10-15	100%	na
6/11	Mild illness hospital worker	>15	55%	na
14/25	Total sensitivity cohort		56%	
12/17	Total sensitivity cohort > 10 days post onset symptoms	>10	71%	
1/13	Healthy blood donors (10), HCoV-OC43 (3)			92%

B. IgM

Positive/ total no. samples	Cohort	Post onset symptoms (days)	Sensitivity	Specificity
3/8	Acute hospitalized patients	<10	38%	na
6/6	Convalescent hospitalized patients	10-15	100%	na
6/11	Mild illness hospital worker	>15	55%	na
15/25	Total sensitivity cohort		60%	
12/17	Total sensitivity cohort > 10 days post onset symptoms	>10	71%	
1/13	Healthy blood donors (10), HCoV-OC43 (3)			92%

2

A. IgG

Positive/ total no. samples	Cohort	Post onset symptoms (days)	Sensitivity	Specificity
3/8	Acute hospitalized patients	<10	38%	na
6/6	Convalescent hospitalized patients	10-15	100%	na
6/11	Mild illness hospital worker	>15	55%	na
15/25	Total sensitivity cohort		60%	
12/17	Total sensitivity cohort > 10 days post onset symptoms	>10	71%	
1/13	Healthy blood donors (10), HCoV-OC43 (3)			92%

B. IgM

Positive/ total no. samples	Cohort	Post onset symptoms (days)	Sensitivity	Specificity
4/8	Acute hospitalized patients	<10	50%	na
6/6	Convalescent hospitalized patients	10-15	100%	na
6/11	Mild illness hospital worker	>15	55%	na
16/25	Total sensitivity cohort		64%	
12/17	Total sensitivity cohort > 10 days post onset symptoms	>10	71%	
1/13	Healthy blood donors (10), HCoV-OC43 (3)	>15		92%

3

A. IgG

Positive/ total no. samples	Cohort	Post onset symptoms (days)	Sensitivity	Specificity
5/8	Acute hospitalized patients	<10	63%	na
6/6	Convalescent hospitalized patients	10-15	100%	na
7/11	Mild illness hospital worker	>15	64%	na
18/25	Total sensitivity cohort		72%	
13/17	Total sensitivity cohort > 10 days post onset symptoms	>10	76%	
1/13	Healthy blood donors (10), HCoV-OC43 (3)			92%

B. IgM

Positive/ total no. samples	Cohort	Post onset symptoms (days)	Sensitivity	Specificity
6/8	Acute hospitalized patients	<10	75%	na
6/6	Convalescent hospitalized patients	10-15	100%	na
7/11	Mild illness hospital worker	>15	64%	na
19/25	Total sensitivity cohort		76%	
13/17	Total sensitivity cohort > 10 days post onset symptoms	>10	76%	
1/13	Healthy blood donors (10), HCoV-OC43 (3)			92%

4

A. IgG

Positive/ total no. samples	Cohort	Post onset symptoms (days)	Sensitivity	Specificity
4/7	Acute hospitalized patients	<10	57%	na
6/6	Convalescent hospitalized patients	10-15	100%	na
6/10	Mild illness hospital worker	>15	60%	na
16/23	Total sensitivity cohort		70%	
12/16	Total sensitivity cohort > 10 days post onset symptoms	>10	75%	
0/25	Healthy blood donors (10), HCoV-OC43 (5), EBV (5), CMV (5)			100%

B. IgM

Positive/ total no. samples	Cohort	Post onset symptoms (days)	Sensitivity	Specificity
4/7	Acute hospitalized patients	<10	57%	na
2/6	Convalescent hospitalized patients	10-15	33%	na
6/10	Mild illness hospital worker	>15	60%	na
12/23	Total sensitivity cohort		52%	
8/16	Total sensitivity cohort > 10 days post onset symptoms	>10	50%	
0/25	Healthy blood donors (10), HCoV-OC43 (5), EBV (5), CMV (5)			100%

5

A. IgG

Positive/ total no. samples	Cohort	Post onset symptoms (days)	Sensitivity	Specificity
4/7	Acute hospitalized patients	<10	57%	na
6/6	Convalescent hospitalized patients	10-15	100%	na
6/10	Mild illness hospital worker	>15	60%	na
16/23	Total sensitivity cohort		70%	
12/16	Total sensitivity cohort > 10 days post onset symptoms	>10	75%	
0/25	Healthy blood donors (10), HCoV-OC43 (5), EBV (5), CMV (5)			100%

B. IgM

Positive/ total no. samples	Cohort	Post onset symptoms (days)	Sensitivity	Specificity
3/7	Acute hospitalized patients	<10	43%	na
1/6	Convalescent hospitalized patients	10-15	17%	na
0/10	Mild illness hospital worker	>15	0%	na
4/23	Total sensitivity cohort		17%	
1/16	Total sensitivity cohort > 10 days post onset symptoms	>10	6%	
0/25	Healthy blood donors (10), HCoV-OC43 (5), EBV (5), CMV (5)			96%

6

A. IgG

Positive/ total no. samples	Cohort	Post onset symptoms (days)	Sensitivity	Specificity
2/7	Acute hospitalized patients	<10	29%	na
6/6	Convalescent hospitalized patients	10-15	100%	na
0/10	Mild illness hospital worker	>15	0%	na
8/23	Total sensitivity cohort		35%	
6/16	Total sensitivity cohort > 10 days post onset symptoms	>10	38%	
0/25	Healthy blood donors (10), HCoV-OC43 (5), EBV (5), CMV (5)			100%

B. IgM

Positive/ total no. samples	Cohort	Post onset symptoms (days)	Sensitivity	Specificity
4/7	Acute hospitalized patients	<10	57%	na
6/6	Convalescent hospitalized patients	10-15	100%	na
2/10	Mild illness hospital worker	>15	20%	na
12/23	Total sensitivity cohort		52%	
8/16	Total sensitivity cohort > 10 days post onset symptoms	>10	50%	
0/25	Healthy blood donors (10), HCoV-OC43 (5), EBV (5), CMV (5)			100%

7

A. IgG

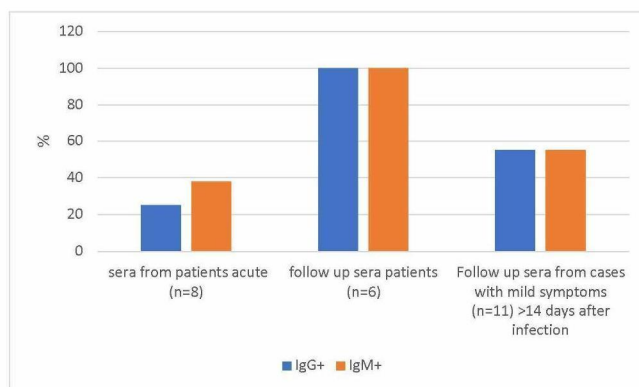
Positive/ total no. samples	Cohort	Post onset symptoms (days)	Sensitivity	Specificity
2/7	Acute hospitalized patients	<10	29%	na
4/6	Convalescent hospitalized patients	10-15	67%	na
0/10	Mild illness hospital worker	>15	0%	na
6/23	Total sensitivity cohort		26%	
4/16	Total sensitivity cohort > 10 days post onset symptoms	>10	25%	
0/25	Healthy blood donors (10), HCoV-OC43 (5), EBV (5), CMV (5)			100%

B. IgM

Positive/ total no. samples	Cohort	Post onset symptoms (days)	Sensitivity	Specificity
0/7	Acute hospitalized patients	<10	0%	na
0/6	Convalescent hospitalized patients	10-15	0%	na
1/10	Mild illness hospital worker	>15	10%	na
1/23	Total sensitivity cohort		4%	
1/16	Total sensitivity cohort > 10 days post onset symptoms	>10	6%	
0/25	Healthy blood donors (10), HCoV-OC43 (5), EBV (5), CMV (5)			100%

Figures 1 to 7 give a graphic representation of the performance of the tests (panels A) including the performance vs the Wantai ELISA (panels B) and virus neutralization test. The panels depict the performance on PCR-confirmed COVID-19 patients

1

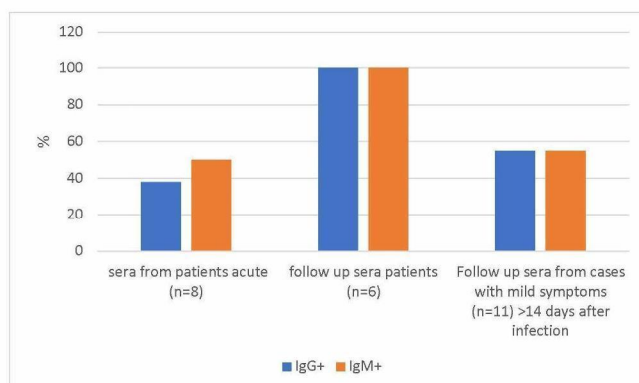


		Wantai ELISA Igtotal		
		low pos*	high pos*	neg
IgG	pos	3	11	
	neg	4	5	2

		Wantai ELISA Igtotal		
		low pos*	high pos*	neg
IgM	pos	3	12	
	neg	4	4	2

**High positives in the Wantai ELISA correlate with detectable titers in the virus neutralization assay. Low positives in the Wantai ELISA did not correlate with detectable presence of neutralizing antibodies.*

2

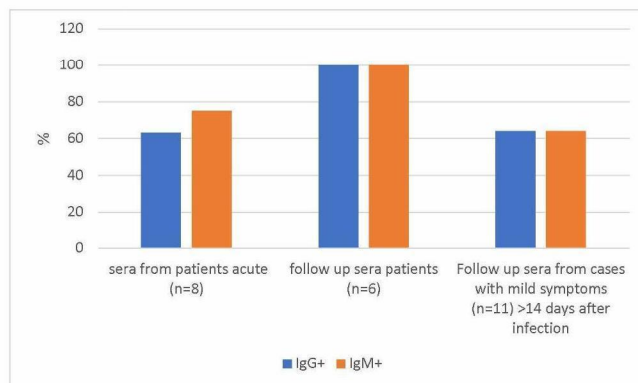


		Wantai ELISA Igtotaal		
		low pos*	high pos*	neg
IgG	pos	3	12	
	neg	4	4	2

		Wantai ELISA Igtotaal		
		low pos*	high pos*	neg
IgM	pos	3	13	
	neg	4	3	2

**High positives in the Wantai ELISA correlate with detectable titers in the virus neutralization assay. Low positives in the Wantai ELISA did not correlate with detectable presence of neutralizing antibodies.*

3

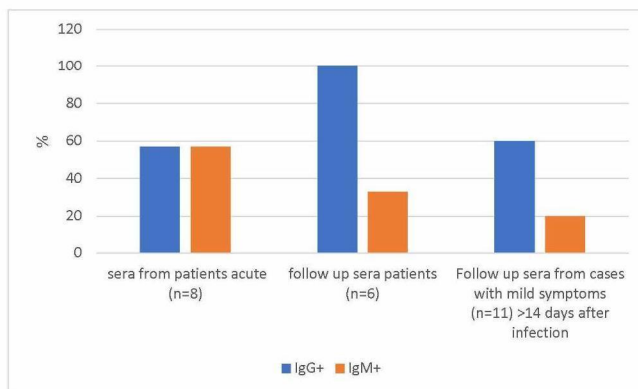


		Wantai ELISA Igtotaal		
		low pos*	high pos*	neg
IgG	pos	4	14	
	neg	3	2	2

		Wantai ELISA Igtotaal		
		low pos*	high pos*	neg
IgM	pos	3	16	
	neg	4		2

**High positives in the Wantai ELISA correlate with detectable titers in the virus neutralization assay. Low positives in the Wantai ELISA did not correlate with detectable presence of neutralizing antibodies.*

4

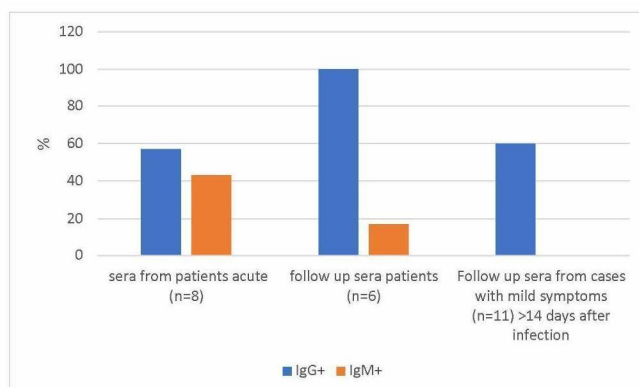


		Wantai ELISA Igtotaal		
		low pos*	high pos*	neg
IgG	pos	3	13	
	neg	3	2	2

		Wantai ELISA Igtotaal		
		low pos*	high pos*	neg
IgM	pos	2	6	
	neg	4	9	2

**High positives in the Wantai ELISA correlate with detectable titers in the virus neutralization assay. Low positives in the Wantai ELISA did not correlate with detectable presence of neutralizing antibodies.*

5

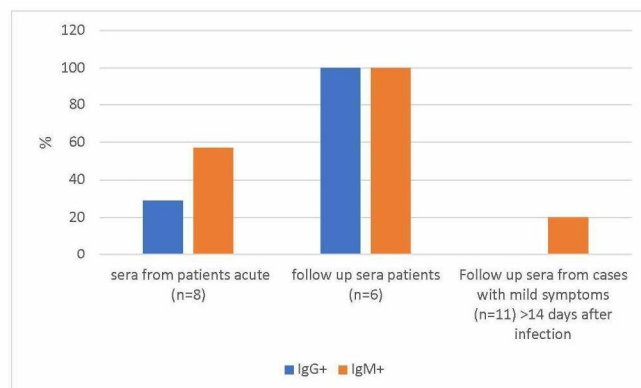


		Wantai ELISA Igtotaal		
		low pos*	high pos*	neg
IgG	pos	3	13	
	neg	3	2	2

		Wantai ELISA Igtotaal		
		low pos*	high pos*	neg
IgM	pos		4	
	neg	6	11	2

**High positives in the Wantai ELISA correlate with detectable titers in the virus neutralization assay. Low positives in the Wantai ELISA did not correlate with detectable presence of neutralizing antibodies.*

6

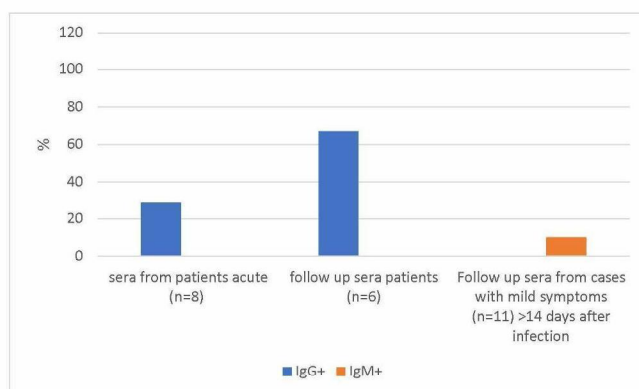


		Wantai ELISA Igtotaal		
		low pos*	high pos*	neg
IgG	pos		8	
	neg	6	7	2

		Wantai ELISA Igtotaal		
		low pos*	high pos*	neg
IgM	pos	1	11	
	neg	5	4	2

**High positives in the Wantai ELISA correlate with detectable titers in the virus neutralization assay. Low positives in the Wantai ELISA did not correlate with detectable presence of neutralizing antibodies.*

7



Wantai ELISA Igtotaal			
IgG	pos	low pos*	high pos*
	neg	6	9
		6	2

Wantai ELISA Igtotaal			
IgM	pos	low pos*	high pos*
	neg	6	14
		1	2

**High positives in the Wantai ELISA correlate with detectable titers in the virus neutralization assay. Low positives in the Wantai ELISA did not correlate with detectable presence of neutralizing antibodies.*

Discussion and conclusion.

Pre-setting the minimal performance required from rapid immuno-assays depends on the use of the rapid test. The seven rapid tests were judged based on the following minimal thresholds (expert opinion):

-individual patient diagnostics: specificity >98%; sensitivity >95% *for both IgG and IgM*

-individual status of having had a SARS-CoV-2 infection in specific (sub)populations, e.g. health care workers and caregivers of vulnerable persons to provide guidance in use of (types of) PPE: specificity >98%; sensitivity >80%. *Only IgG.*

-sero-epidemiological studies (e.g. collecting seroprevalence data as proxy for herd immunity, input in models): specificity >98%; sensitivity >90%. *Only IgG.*

Based on the first limited validation presented here, it can be concluded that based on the overall data for samples taken > 10 days post onset symptoms for PCR-confirmed COVID-19 cases none of the seven tests fulfilled the sensitivity criteria for IgG set above. When analyzing with sera taken 10-15 days post onset of symptoms from hospitalized patients, all tests but one had a sensitivity of 100% for IgG. However, when analyzing with sera from mild patients taken > 15 days post onset of symptoms, a wide range of sensitivities for IgG were observed, varying from 0%-64%. In these cases none of the tests fulfilled the preset criteria. It should be noted that PCR was the reference test here.

Looking at specificity, 3, 1, and 2 tests did not reach the preset threshold for specificity of IgG. However, the result of 92% specificity was obtained as one of 13 samples in the specificity panel gave a false-positive result. To determine more precisely the specificity of these and the other four tests it is absolutely required that a larger specificity panel is tested.

When comparing the IgG performance of the rapid tests vs the Wantai test and specifically those sera that were positive in both the Wantai and the neutralization test (indicated as high pos for the Wantai test), it was observed that the 3 test showed the best performance with 14 of 16 (87.5%) samples with a neutralization titer found positive. This was closely followed by 5 and 4 (both 13 of 15; 86.7%), 2 (12 of 16; 75%) and 1 (11 of 16; 68.7%).

None of the seven tests fulfilled the preset minimal requirements for test specificity and sensitivity for the three different operational contexts that were defined. These data underline the importance of extensive validation in the right (sub)populations and settings to avoid guidance of control efforts at individual and population level based on false diagnosis of individuals. Until extensive validation gives evidence that the accuracy of the tests is high in specific populations and settings, it is not appropriate to use rapid immuno-assays for clinical decision making, to guide dedicated measures for specific subpopulation and to guide general control measures.

VS 2.0 15 April 2020,